Extrapulmonary *Pneumocystis Jirovecii* Presenting as Fever and Lymphadenopathies. Case Report and Literature Review

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**Abstract**

Pneumocystosis is one of the most common opportunistic infection occurring in HIV infected patients. The usual clinical presentation of Pneumocystis infection is pulmonary. Extrapulmonary Pneumocystis (EP) infections are uncommon, and have generally been observed in patients with advanced HIV disease, many times under second line prophylaxis with aerosolized pentamidine. Because of its low frequency, its low index of suspicion and a variety of clinical presentations, extrapulmonary forms of *P. jirovecii* infections are usually difficult to diagnose. We report a case of extrapulmonary pneumocystosis as a form of presentation of Acquired Immunodeficiency Syndrome (AIDS) presenting with fever and enlarged intra-abdominal lymph nodes, and review all published cases of extrapulmonary pneumocystosis from 1997 to 2017.

**Keywords:** *Pneumocystis Jirovecii*; AIDS; HIV; Extrapulmonary Infection; Lymphadenopathy

**Introduction**

Symptomatic infection caused by *Pneumocystis Jirovecii* is an opportunistic infection that occurs almost exclusively in immunocompromised hosts. Due to the microorganism’s low intrinsic virulence, infection in immunocompetent host is usually subclinical. Pneumocystosis is the most common opportunistic
Computed tomography (CT) of thorax, abdomen and pelvis with intravenous contrast were obtained (Figure 1), showing spleen and liver enlargement with homogeneous density. Enlarged lymph nodes were noted at the retroperitoneum, para-aortic space, portacaval space, retro pancreatic space and at the mesentery root, with extension up to the iliac arteries bifurcation. Thorax CT showed no abnormalities. Showing spleen and liver enlargement with homogeneous density. Enlarged lymph nodes were noted in the retroperitoneum, para-aortic space, portacaval space, retropancreatic space and at the mesentery root, with extension up to the iliac arteries bifurcation. Chest CT showed no abnormalities. Mycobacterial and fungal blood cultures were negative. Other serological and microbiological tests are presented in Table 1.

A bone marrow biopsy and a laparoscopic biopsy of retroperitoneal lymph nodes were obtained, and sent for pathological examination and flow cytometry analysis. Samples for mycobacterial, fungal and usual microorganism cultures were also collected. Antimicrobial prophylaxis with azithromycin and trimethoprim-sulfamethoxazole (TMP-SMX) was started. Because a lymphoproliferative process was suspected, prednisone 40 mg/day was started. On June 23rd, results were received for absolute CD4+ count (35/mm³) and HIV viral load (298,000 copies/mL).

<table>
<thead>
<tr>
<th>Test</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>EBV-IgG</td>
<td>Positive</td>
</tr>
<tr>
<td>EBV-IgM</td>
<td>Negative</td>
</tr>
<tr>
<td>EBNA</td>
<td>Negative</td>
</tr>
<tr>
<td>EBV-EA</td>
<td>Negative</td>
</tr>
<tr>
<td>Viral load EBV</td>
<td>Non-detectable</td>
</tr>
<tr>
<td>CMV-IgG</td>
<td>Positive</td>
</tr>
<tr>
<td>CMV-IgM</td>
<td>Negative</td>
</tr>
<tr>
<td>Viral load CMV</td>
<td>Non-detectable</td>
</tr>
<tr>
<td>Toxoplasmosis serology</td>
<td>Non-reactive</td>
</tr>
<tr>
<td>Chagas disease serology</td>
<td>Non-reactive</td>
</tr>
<tr>
<td>PVB19 IgG</td>
<td>Negative</td>
</tr>
<tr>
<td>VDRL</td>
<td>Negative</td>
</tr>
<tr>
<td>T. pallidum-HA assay</td>
<td>Negative</td>
</tr>
<tr>
<td>FTAbs</td>
<td>+1/4 positive.</td>
</tr>
<tr>
<td>C. neoformans antigenemia</td>
<td>Negative.</td>
</tr>
<tr>
<td>H. Capsulatum antibodies</td>
<td>Negative.</td>
</tr>
<tr>
<td>HBSAg and antibodies to HCV</td>
<td>Negative.</td>
</tr>
</tbody>
</table>

Table 1: Microbiological and serological tests results. EBV: Epstein Barr virus; EBNA: Epstein Barr Nuclear Antigen; EA: early antigen; CMV: cytomegalovirus; PV: Parvovirus.

Figure 2(a,b): PAS and Grocott stains shows numerous elements, 5-8 microns large, mainly extracellular, intensively PAS-positive, with a rounded or oval morphology and a PAS positive-central area, resembling “crushed ping pong balls”, a morphology corresponding to *P. jirovecii* cysts.
Antiretroviral drugs (tenofovir, lamivudine and raltegravir) were requested to start treatment. Bone marrow and lymph node cultures yielded no microorganism growth.

Flow cytometry analysis and pathology of the bone marrow showed a marked inversion of CD4 to CD8 ratio, with no phenotypic anomalies, no evidence of lymphoproliferative disease, and with marked reactive changes.

On July 1st, the initial report on the retroperitoneal lymph node informed atypical lymphoid hyperplasia, "probably related to viral infection". Later, special stains (PAS and Grocott) revealed numerous elements, 5-8 microns large, mainly extracellular, intensely PAS-positive, with a rounded or oval morphology and a PAS positive-central area, resembling "crushed ping pong balls", a morphology corresponding to *P. jirovecii* cysts (Figure 2). Immunofluorescent staining with specific monoclonal antibodies confirmed the diagnosis.

Treatment with intravenous TMP-SMX (15 mg/kg/day trimethoprim) was then started. The patient’s clinical condition rapidly improved, becoming afebrile on day 6 after starting treatment. On day 13, TMP-SMX was changed to oral administration, and the patient was discharged. He completed 21 days of TMP-SMX. The patient fully recovered from his infection, but he was lost to follow-up when he returned to his home country (Brazil) 2 months after discharge.

**Discussion**

*Pneumocystis Jirovecii* is one of the most important opportunistic pathogen affecting AIDS patients. Incidence of *P. jirovecii* infection (which before the availability of highly active antiretroviral treatment, (HAART), affected up to 75% of HIV infected patients) is around 3.9 /1000 year-patient [2], and it is still the most common opportunistic infection in this patient group, especially in those with low CD4 count or in patients unaware of their HIV infection.

Although it has been formerly classified as a protozoa, *P. jirovecii* is a yeast-like fungus, classified as such on the basis of genomic sequences of its ribosomal RNA, its wall chemical composition, and the structure of key enzymes [3]. However, Pneumocystis is an atypical fungus that does not grow in culture media.

Human Pneumocystis is an anthropozoonosis, being humans the only reservoir and host. Transmission is primarily airborne (usually from other infected persons, and in certain occasions, possibly from environmental reservoirs) [4]. Colonization with *P. jirovecii* occurs in the early stages of life, but in the presence of a normal immunologic response, the infection is contained, although not eradicated. Infection persists in a dormant state, with the potential of progression to clinical disease if there is a derangement of the host’s immunological response. Rarely, a recently acquired infection may progress to pulmonary and eventually extrapulmonary clinical disease. From the lung, the microorganism may spread systemically either directly or by lymphatic or hematogenous pathways, although dissemination is distinctly unusual.

Pulmonary pneumocystosis is usually an interstitial pneumonia with symptoms as fever, cough and dyspnea. Radiological appearance is customarily a diffuse reticular or finely granular interstitial infiltrate. On CT, bilateral patchy “ground glass" asymmetric infiltrates, with upper lobes predominance is usually observed [5]. Occasionally, lung pneumocystosis may occur in the absence of abnormal CT findings.

Although pulmonary pneumocystosis is the most prevalent serious lung infection in HIV infected patients, extrapulmonary pneumocystosis is uncommon.

*P. jirovecii* has the potential to disseminate from the lung to other organs, producing infection in almost any anatomic site. Endothelial damage due to Kaposi’s sarcoma, cytomegalovirus infection, or recurrent *P. jirovecii* infection may facilitate dissemination from the lung.

Before HIV epidemic, extrapulmonary pneumocystosis was extremely rare (only 16 cases reported from 1954 to 1996, mainly post-mortem studies). HIV epidemic was accompanied by an increase in Pneumocystis cases, and by an increase in extrapulmonary pneumocystosis reports. The true incidence of extrapulmonary pneumocystosis is unknown, although it has been estimated in 0.06-2.5% [6-8].

Pulmonary infection and extrapulmonary infection have been considered to be caused by different genetic variants of *P. jirovecii*, variants that may potentially differ in their susceptibility to antimicrobial treatments [9].

Extrapulmonary pneumocystosis has been observed in HIV infected patients receiving aerosolized pentamidine prophylaxis. The distribution of this drug within the lung, when administered this way, is not uniform [10] and it
has no systemic effects. Therefore, it may be insufficient to prevent systemic dissemination and to suppress *P. jirovecii* out of the lung, which explains its association as a risk factor for extrapulmonary pneumocystosis [11,6].

Other risk factors observed are advanced stages of HIV infection with no prophylaxis - in these patients, extrapulmonary pneumocystosis without simultaneous lung disease may be observed-, low CD4 count and a history of lung infections.

Extrapulmonary pneumocystosis has been described in eyes (usually affecting the choroidal layer), ear, thyroid, spleen and many other locations [12].

Cases published as of 1997 have been thoroughly reviewed by Ng, et al. The epidemiology of extrapulmonary pneumocystosis may have changed after that, as the availability and tolerability of HAART increased along the years. We therefore searched for cases (either in English or Spanish, in PubMed and Scielo databases) not included in Ng review, published from 1997 to 2017.

Lymph nodes are the most common site for extrapulmonary *P. jirovecii* infection in HIV positive patients. Among the 90 cases (published between 1982 and 1996) reviewed by Ng, et al. 19 (21%) had lymph node involvement13.

In a review of 233 autopsy findings (HIV positive patients died between from 1985 to 1996), pulmonary pneumocystosis was observed in 56 patients. Within this group, extrapulmonary involvement was observed in 7 (13%), six of them with lymph node disease due to *P. jirovecii* [13]. Intrathoracic adenopathies have been documented in 18% of chest CTs in patients with pulmonary pneumocystosis, although as a rule, lymph node involvement is unapparent in usual chest x-ray [14].

It is possible that the clinical behavior of *P. jirovecii* may have varied according with the increased availability of better antiretroviral treatments. Initial observations include patients from a time in which many of the currently used antiretroviral drugs were not still available (by 1994, the only approved drugs were zidovudine, didanosine, zalcitabine and stavudine). In the reviewed literature (published in English and Spanish, from 1997 to 2017), among 34 cases of extrapulmonary pneumocystosis, 10 cases presented documented or potential lymph node involvement caused by *P. jirovecii*. In 6 post-mortem cases, a detailed description was not provided [13]. The four remaining cases comprised one patient with pulmonary pneumocystosis who developed hilar adenopathies and pleural masses while on dapsone prophylaxis (not biopsied, but resolved with anti-Pneumocystis treatment) [15]; a 7-year-old boy with generalized lymphadenopathies, in whom fine needle aspiration cytology allowed diagnosis [16]; one patient with mediastinal, cervical, and left supraclavicular lymphadenopathies (surgical biopsy positive for Pneumocystis) [17] and one patient with simultaneous documented involvement of lung and colon, with potential infection of mesenteric and retroperitoneal lymph nodes (not biopsied) [18].

Relevant clinical data from extrapulmonary Pneumocystis cases published from 1997 to 2017 is summarized in Table 2 [13,15-32].

Additionally, one 50-year-old male patient was reported in an Italian journal in 1998. He was diagnosed with AIDS 8 months before, treated with zidovudine and didanosine, with an absolute CD4 count 6 /mm3. He developed a severe respiratory pneumocystosis (in spite of being on aerosolized pentamidine prophylaxis) and was treated with pirimetamine and clindamycin. He died on day 14 after diagnosis. On autopsy, a massive pulmonary embolism was observed. Grocott stain allowed to document persistence of pulmonary infection and hilar and mediastinal lymphatic dissemination of Pneumocystis [33].

Because Pneumocystis cannot be readily cultured in the routine laboratory, the gold standard for diagnosis is the direct visualization of the organisms. Extrapulmonary *P. jirovecii* infection is diagnosed by the observation of the typical cyst in formalin-fixed tissue. It should be noted that *P. jirovecii* is usually very difficult to identify with hematotoxilin-eosin or Papanicolaou stains, and they stain faint and variably. Grocott stain (silver methenamine) allow the organism to be seen mainly as spherical cysts, with variations in shape. Occasionally, small interior structures can be observed in the form of commas, dots, or rings. Differential diagnosis include organisms similar to *P. jirovecii*, such as Histoplasma capsulatum, capsule-deficient Cryptococcus, Penicillium marneffei, endospores of Coccidioides spp and Candida species, but these spores usually have a budding appearance with is absent in *P. jirovecii* cysts [16,34]. Direct immunofixation or PCR techniques can be used for diagnosis in difficult cases.

Sometimes, (as when disease is localized in the eye) diagnosis is based on typical appearance of lesions and response to treatment. Prognosis is better for localized
forms in eye or ear than for disseminated forms affecting non-contiguous sites [12].

The differential diagnosis of lymphadenopathies in patients with HIV infection includes HIV associated-lymphadenopathy, neoplastic diseases such as Kaposi sarcoma or lymphoproliferative diseases, and a variety of infectious etiologies such as mycobacterial infection (tuberculous and non-tuberculous) and fungal infections. Lymph node disease is a frequent diagnostic challenge for physicians who treat HIV infected patients, as up to 25% of HIV infected patients experience some form of lymph node involvement, at any point of the clinical progression of the disease, from early stages to advanced HIV infection. Our observation emphasizes that, although highly uncommon, extrapulmonary pneumocystosis may be considered in the differential diagnosis of lymph node disease.

Treatment of extrapulmonary pneumocystosis follows the same principles of pulmonary Pneumocystis treatment. Trimethoprim-sulfamethoxazole (TMP-SMX) remains the drug of choice for all forms of Pneumocystis infection. The recommended daily dose is TMP 15-20 mg/kg plus SMX 75-100 mg/kg. The optimal treatment duration of extrapulmonary Pneumocystis has not been well studied, but it is generally recommended to treat HIV-infected patients for at least 21 days [35]. Second line therapy includes intravenous pentamidine, clindamycin plus primaquine, atovaquone, and dapsone plus TMP22. Caspofungin associated with clindamycin [36] or low doses SMX-TMP [37] may also be considered among the therapeutic alternatives.

**Conclusions**

The present case illustrates the importance of considering Pneumocystis in the differential diagnosis of lymphadenopathies in AIDS patients. Although uncommon, even in this era of highly active antiretroviral treatments, extrapulmonary *P. jirovecii* infections may still occur. Knowledge of the risk factors for extrapulmonary pneumocystosis (such as HIV infection advanced stage, with profound immunosuppression, or use of aerosolized pentamidine prophylaxis) and of the variety of extrapulmonary *P. jirovecii* clinical forms may allow for timely diagnosis and appropriate management.

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